

REVIEW

The role of earthworm defense mechanisms in ecotoxicity studies**R Roubalová, P Procházková, J Dvořák, F Škanta, M Bilej***Laboratory of Cellular and Molecular Immunology, Institute of Microbiology of the Academy of Sciences of the Czech Republic, v. v. i., Vídeňská 1083, 142 20, Prague 4, Czech Republic*

Accepted July 24, 2015

Abstract

Earthworms are important soil organisms that affect the soil structure by influencing organic and inorganic matter breakdown. Earthworms are in permanent contact with soil particles via their permeable skin and digestive tract and are thus strongly affected by pollutants present in the soil. Earthworms often live in very hostile environments with an abundant microflora and therefore have developed very potent defense mechanisms. These mechanisms have been described to be influenced by various types of organic and inorganic pollutants and also by the nanoparticles that reach the soil system. Reduced abilities of earthworms to protect themselves against pathogenic microorganisms result in lower reproduction rates and increased mortality. In this review, a summary of the up-to-date data describing the effects of contaminants on the natural defense barriers and immune system of earthworms is presented.

Key Words: pollution; immune system; earthworms; biomarker**Introduction**

Earthworms (Lumbricidae, Annelida) are protostomian organisms with a true celom that is filled with celomic fluid containing free celomocytes. The celomic cavity is metameric, and the segments are separated by transversal septa. Each segment of the cavity interfaces with the outer environment via a pair of metanephridia and a dorsal pore that enables microorganisms to enter the celomic cavity. Therefore, the celomic fluid is not aseptic and contains bacteria, fungi and protozoa from the outer environment. The growth of these microorganisms is kept under control by various cellular and humoral innate defense mechanisms that will be described in detail in the following section.

Earthworms are the most abundant invertebrates in the soils of temperate regions and are extremely important for soil formation (Edwards, 2004). Earthworms participate in nutrient cycling in terrestrial ecosystems and in the formation of the soil profile from the physical, chemical and microbial perspectives (Bartlett *et al.*, 2010). They improve its structure by increasing the macroporosity, which

affects aeration, water dynamics and organic and inorganic matter breakdown (Wen *et al.*, 2006; Ruiz *et al.*, 2011). Earthworms are permanently in close contact with soil particles and microorganisms present in the soil via both a highly permeable skin and an alimentary tract (Jager *et al.*, 2003; Drake and Horn, 2007). Therefore, they are significantly affected by the pollutants that reach the soil system and are thus well suited for the monitoring of soil contamination. Different earthworm species have different effects on soil formation because of their different behavioral patterns. Epigeic earthworms live above the mineral soil, rarely form burrows and preferentially feed on plant litter. Endogeic species live below the surface, where they build predominantly horizontal burrows. These species ingest large amounts of mineral soils and humified material. Anecic earthworms build permanent vertical burrows deep into the mineral soil layer and come to the surface to feed on decomposed plant litter and other organic residues (Lee, 1985). Two epigeic species, *i.e.*, *Eisenia fetida* and *Eisenia andrei*, have been used for many years to monitor ecotoxicity. There are two sets of guidelines, *i.e.*, those from the Organization for Economic Co-operation and Development (OECD) and those from the International Organization for Standardization (ISO), for the assessment of the ecological risk of contaminated soil, the determination of the acute toxicity of chemicals on earthworms (OECD, 1984;

Corresponding author:

Radka Roubalová
Laboratory of Cellular and Molecular Immunology
Institute of Microbiology of the Academy of Sciences of the
Czech Republic, v. v. i.
Vídeňská 1083, 142 20, Prague 4, Czech Republic
E-mail: r.roubalova@biomed.cas.cz

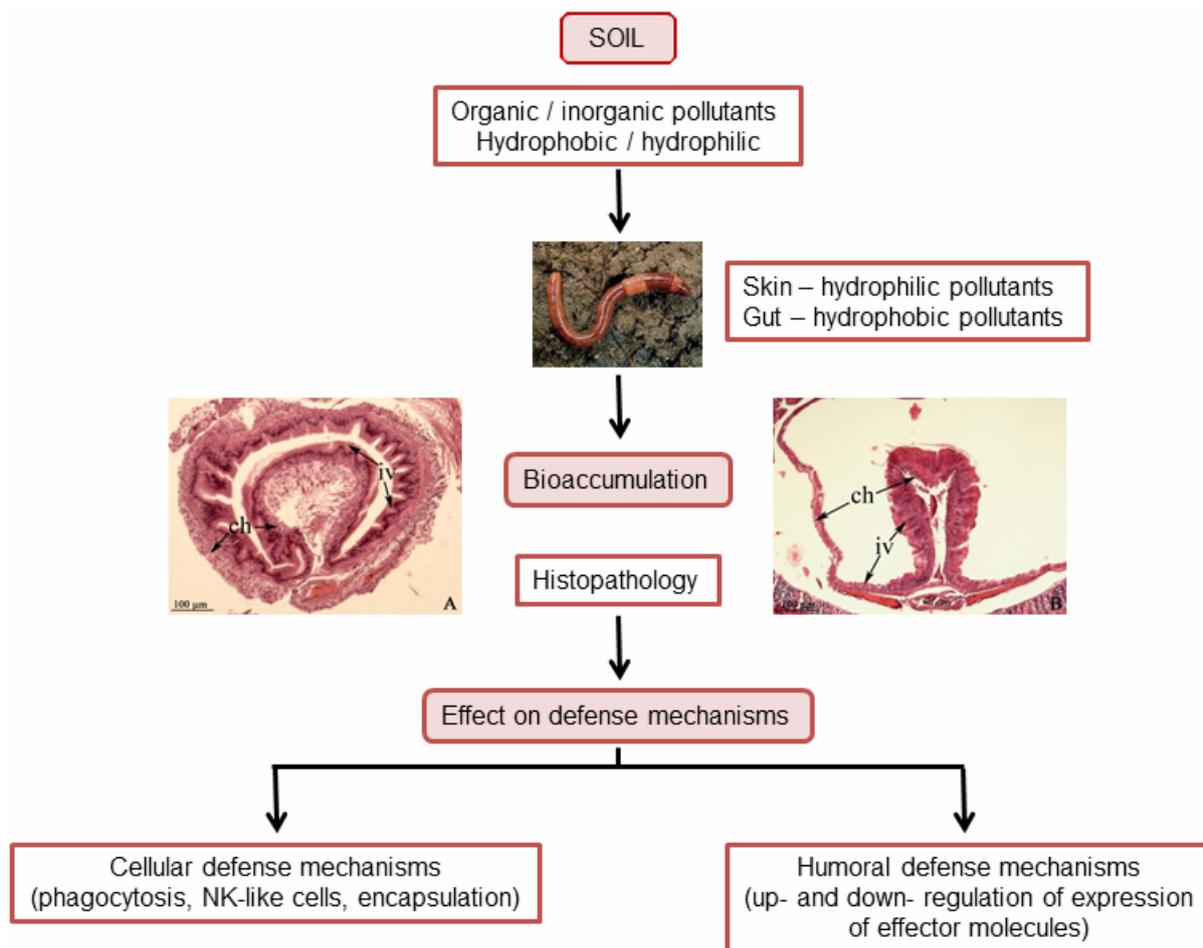


Fig. 1 Earthworms are affected by the presence of pollutants in the soil. Hydrophilic contaminants enter the earthworm body predominantly through the skin, whereas hydrophobic substances enter via the digestive tract. Pollutants are accumulated in earthworm tissues, which can result in tissue and cell disruption, such as progressive reduction of intestinal villi (iv) and chloragogenous tissue (ch) in earthworms kept in dioxin-polluted soil (B; Roubalova *et al.*, 2014). Additionally, both cellular and humoral defense mechanisms are impaired by the soil contaminants.

ISO, 1993), and the effect on their reproduction (ISO, 1998; OECD, 2004).

Earthworms have been described to bioaccumulate contaminants, such as various organic pollutants (Jager *et al.*, 2005), heavy metals (Nahmani *et al.*, 2007) and nanoparticles (Canesi and Prochazkova, 2014). They are able to take up chemicals from pore water through their skin and via soil ingestion. According to the model developed by Belfroid *et al.* (1995), the ingestion of sediment can be the dominant uptake route of hydrophobic compounds with $\log K_{ow}$ values > 5. The presence of contaminants in the soil disturbs major physiological functions of earthworms, such as survival, nutrition, immunity, growth, and reproduction, and these effects depend on the matrix, exposure time, and the types and doses of the pollutants in the environment. In recent years, there has been a growing interest in increasing our knowledge of the

biological responses of earthworms to pollutants in order to standardize a suite of biomarkers of the responses to soil chemical pollution (Beliaeff and Burgeot, 2002). Biomarkers detect the effects of contamination at an early stage before sublethal effects, such as inhibition of growth and reproduction, become apparent. The biomarker approach represents a very useful tool in monitoring stress response to pollutants in field populations (Kammenga *et al.*, 2000; Hankard *et al.*, 2004). The choice of appropriate biomarkers is crucial for monitoring the effects of pollution on organisms. Reactions to pollution may be monitored on various levels, the whole body level (viability, weight loss, reduction of reproduction, and escape reaction), the organ and tissue level (histopathological changes), the cellular level (decrease in the physiological conditions of the cells) and the molecular level (the up- and down-regulation of the expression levels of

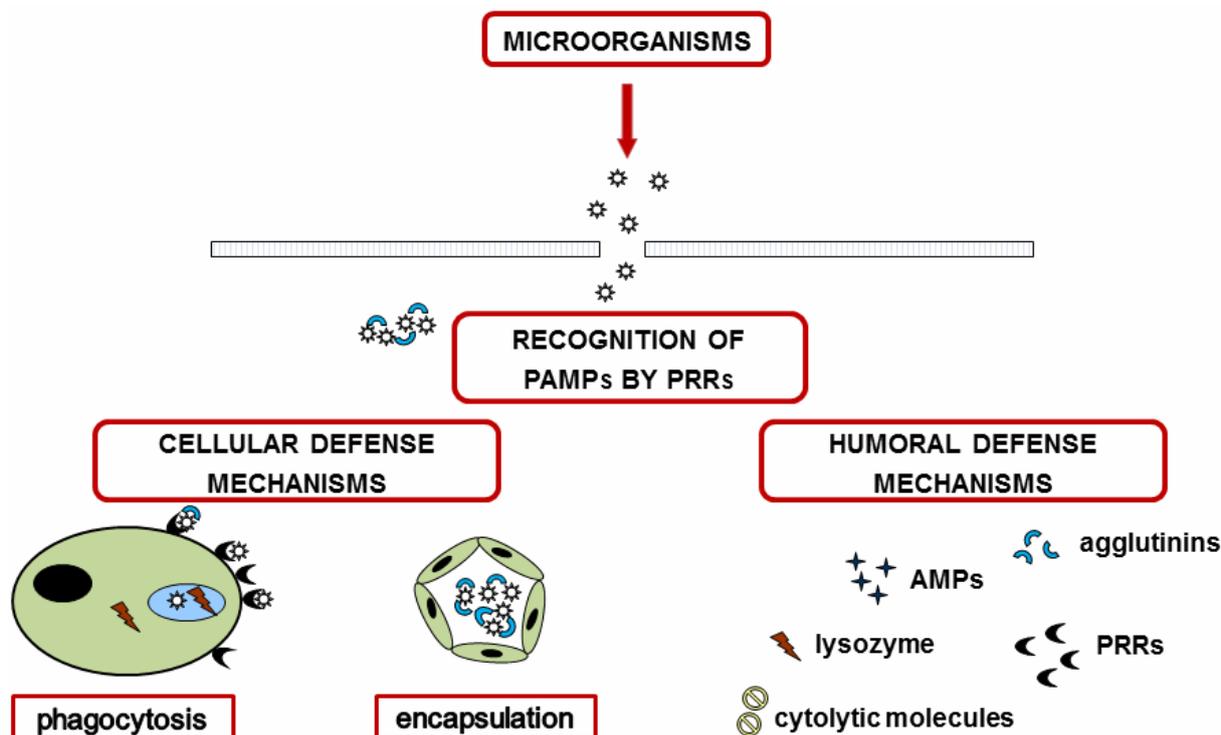


Fig. 2 The general scheme of the innate defense mechanisms in earthworms. The first protective barrier of earthworms is the skin in combination with the secreted mucus that contains various antimicrobial factors. Invading microorganisms are recognized by both soluble and membrane-bound pattern recognition receptors (PRRs) that sense pathogen-associated molecular patterns (PAMPs). On the basis of this recognition, microorganisms are phagocytized by coelomocytes or agglutinated and subsequently encapsulated. Moreover, genes encoding various humoral factors involved in the elimination of invaders are expressed, such as antimicrobial peptides (AMPs), cytolytic molecules, agglutinins, lysozyme and various soluble PRRs that trigger the activation of the prophenoloxidase cascade.

genes that are sensitive to the environmental changes, transcriptome profiling) (Owen *et al.*, 2008; Asensio *et al.*, 2013; Calisi *et al.*, 2013; Roubalova *et al.*, 2014; Sanchez-Hernandez *et al.*, 2014; Sforzini *et al.*, 2015) (Fig. 1). Although these responses may indicate the disturbances at the level of populations, only few data link biomarker level with effects on the functioning of earthworms in ecosystems (Maboeta *et al.*, 2003; Spurgeon *et al.*, 2005; Plytycz *et al.*, 2009).

The effects of pollutants on the defense mechanisms of earthworms

Similarly to other invertebrates, earthworms rely on natural nonspecific innate immunity for defense and lack anticipatory, specific and lymphocyte-based immune mechanisms. Additionally, the natural barriers of earthworms represent the first line of protection against the invasion of microorganisms. A brief summary of earthworm immune mechanisms is shown in Figure 2. In the following sections, the effects of various soil pollutants on the nonspecific defense barriers and the cellular and humoral mechanisms of immunity are reviewed.

Natural defense mechanisms and pollution

The first nonspecific protective barrier in earthworms is the skin, which consists of the epidermis and a thin cuticle that covers the entire body. The epidermis is formed by a single-layer epithelium of supporting cells, basal cells that have an important role in wound healing and graft rejection, and secretory cells that secrete mucus containing mucopolysaccharide-lipid-protein complex (Alves *et al.*, 1984; Bernaldo de Quiros and Benito, 1986) that serves as a lubricant during locomotion and contains several antimicrobial factors (Valembouis *et al.*, 1986). The cuticle contains mucopolysaccharides that act as an antimicrobial barrier (Rahemtulla and Lovtrup, 1974).

Both cuticle and mucus production can be affected by the inorganic and organic contaminants as well as nanoparticles present in the soil. The exposure of the earthworms *Lumbricus rubellus* and *Lumbricus variegatus* to C60 fullerene nanoparticles has been described to result in cuticle damage with underlying pathologies of the epidermis and muscles (Pakarinen *et al.*, 2011; Van Der Ploeg *et al.*, 2013). Furthermore, the exposure of *E. fetida* to sub-lethal concentrations of 1,2,4-trichlorobenzene

Table 1 Summary of recent studies involving genotoxicity assessment of various organic and inorganic pollutants

Tested species	Organic pollutant	Source	Reference
<i>E. fetida</i>	naphthenic acids	constituents of petroleum, used in commercial and industrial applications	(Wang <i>et al.</i> , 2015a)
<i>E. fetida</i>	di-n-butyl phthalates	increase the plasticity of many materials	(Du <i>et al.</i> , 2015)
<i>E. fetida</i>	benzo[a]pyrene	the result of incomplete combustion	(Duan <i>et al.</i> , 2015)
<i>E. fetida</i>	triclosan	antimicrobial additive used in personal care products	(Lin <i>et al.</i> , 2014)
<i>E. fetida</i>	metalaxy-M	fungicide	(Liu <i>et al.</i> 2014)
<i>E. fetida</i>	azoxystrobin	fungicide	(Han <i>et al.</i> , 2014)
<i>E. fetida</i>	chlortetracycline	veterinary antibiotics	(Lin <i>et al.</i> , 2012)
<i>E. andrei</i>	B[a]P, TCDD	by-products from a number of human activities	(Sforzini <i>et al.</i> , 2012)
<i>E. fetida</i>	toluene, ethylbenzene and xylene	associated with crude petroleum and petroleum products	(Liu <i>et al.</i> , 2010)
Tested species	Inorganic pollutant	Source	Reference
<i>E. andrei</i>	Cd, Zn	metals provided in the form of CdSO ₄ , ZnSO ₄	(Otomo <i>et al.</i> , 2014)
<i>E. fetida</i>	Cr, Cu, Ni, Pb, Zn	soils subjected to chemical characterization and total main heavy metal quantification	(Zheng <i>et al.</i> , 2013)
<i>L. castaneus</i> , <i>D. rubidus</i>	As	concentrations of arsenic elevated due to mining	(Button <i>et al.</i> , 2012)
<i>A. caliginosa</i> , <i>E. fetida</i>	Cu, Cd	sites near roads with heavy traffic	(Klobucar <i>et al.</i> , 2011)
<i>E. andrei</i>	Be, Al, Ba, Mn, Fe, Ni, Zn, U	deposition of mine tailings and sludge, runoffs from the aquatic system	(Lourenco <i>et al.</i> 2011)
<i>E. fetida</i>	Ni, Cr(III), Cr(VI)	pollutants used in numerous industrial processes	(Bigorgne <i>et al.</i> , 2010)
<i>E. fetida</i>	Cd, Pb	toxic elements widely distributed in the environment	(Li <i>et al.</i> , 2009)

results in ultrastructure alterations of the cuticle and skin, and the reduction of mucus production by secretory cells. At higher concentrations, mucus production disappears, and the cuticle is loosened and weakened (Wu *et al.*, 2012a). Exposure of the earthworm *E. fetida* to soil containing tetraethyl lead (TEL) and lead oxide (a gasoline additive) causes ruptures of the cuticle and skin, extrusion of the coelomic fluid and inflexible metameric segmentation (Venkateswara Rao *et al.*, 2003).

Cellular innate immunity

The celomic fluid of earthworms contains different types of cells that are generally termed celomocytes. The nomenclature of celomocytes is based on differential staining, ultrastructure, and granular composition. There are two basic categories of celomocytes. Amebocytes function primarily in immune reactions, such as phagocytosis, encapsulation, nodulation as well as humoral immune responses, and mainly nutritive

eleocytes (Sima, 1994). Celomocytes have been described to respond to a wide range of pollutants and therefore are often used in soil ecotoxicology assessment.

At the cellular level, two immune system-related parameters have been used as sensitive sub-lethal endpoints in assessment of the toxicity of pollutants in earthworms: phagocytosis and NK-like cell activity. Phagocytosis represents an important defense mechanism that begins with the recognition of non-self, which is followed by the engulfment and destruction of phagocytosed particles. Engulfed material can be eliminated by proteolytic and lysosomal enzymes or by an oxidative burst that involves the production of highly reactive oxygen radicals. The inhibition of phagocytosis in earthworms that are exposed to various metals and organic substances, such as polychlorinated biphenyls (PCBs) and polychlorinated dibenzo-p-dioxins/dibenzofurans (PCDDs/Fs), has been described (Ville *et al.*, 1995; Fugere *et al.*, 1996; Fournier *et al.*, 2000; Sauve *et al.*, 2002; Belmeskine *et al.*, 2012). Silver nanoparticles have been shown to be accumulated predominantly in the amoebocyte population of celomocytes with subsequent selective cytotoxicity of these cells (Hayashi *et al.* 2012). Furthermore, some celomocytes have been shown to possess cytotoxic activity similar to that of natural killer (NK) cells. These cells exhibit rapid response to allogenic structures and have been described to be involved in the rejection of allografts (Suzuki and Cooper, 1995). The NK-like cell activity has been demonstrated to be suppressed by polyaromatic hydrophobic hydrocarbons (PAHs) (Patel *et al.*, 2007), PCBs (Suzuki *et al.*, 1995), and PCDDs/Fs (Belmeskine *et al.*, 2012). Furthermore, flow cytometry has revealed a lower frequency of immune cells (amoebocytes) in contrast with metabolic eleocytes in earthworms that have been exposed to metal- and radionuclides-contaminated soil (Lourenco *et al.*, 2011).

At the subcellular level, the lysosomal membrane stability system has been identified as a specific target of the toxic effects of contaminants (Moore, 1990). Lysosomal membrane integrity can be measured with the neutral red retention assay (Weeks and Svendsen, 1996). The stability of the membranes has been shown to decrease with increasing stress due to the presence of pollutants in the environment (Moore, 1985; Booth and O'Halloran, 2001; Booth *et al.*, 2003).

Because many soil contaminants exert genotoxic activities that result in DNA damage in the celomocytes, it is used as an important tool in environmental biomonitoring. The most widely used genotoxicity biomarker is the comet assay; this method has been shown to be appropriate for measuring DNA damage in the individual cells of both vertebrates and invertebrates (Singh *et al.*, 1988; Fairbairn *et al.*, 1995; Cotelle and Ferard, 1999; Faust *et al.*, 2004; Sforzini *et al.*, 2012). In Table 1, examples of organic and inorganic pollutants described in recent studies that cause DNA damage are listed.

Humoral defense mechanisms

Molecules involved in innate immunity

The celomic fluid of annelids exhibits numerous biological activities that are involved in the defense mechanisms against invaders (Fig. 2). The recognition of microbial pathogens is mediated by pattern recognition receptors (PRRs) that sense so-called pathogen-associated molecular patterns (PAMPs). These structures are common among microorganisms and include, *i.e.*, the lipopolysaccharides of Gram-negative bacteria, constituents of the peptidoglycan of Gram-positive bacteria, β -glucans of yeasts and viral double-stranded RNA. This recognition results in the activation of both cellular and humoral defense mechanisms, including the production of antimicrobial proteins and peptides (Joskova *et al.*, 2009), and the activation of an important invertebrate defense mechanism termed the prophenoloxidase cascade (Beschin *et al.*, 1998; Soderhall and Cerenius, 1998).

To date, only two PRRs in earthworms have been described, *i.e.*, celomic cytolytic factor (CCF) (Beschin *et al.*, 1998; Bilej *et al.*, 1998, 2001) and Toll-like receptor (TLR) (Skanta *et al.*, 2013), and these PRRs recognize various PAMPs. The expression of CCF has been described to be significantly down-regulated in *L. rubellus* following lifelong exposure to C60 nanoparticles, which suggests the induction of immunosuppression (Van Der Ploeg *et al.*, 2013). Dioxins have also been shown to affect the expression of CCF (Roubalova *et al.*, 2014).

A wide range of antimicrobial molecules that are involved in killing the microorganisms that enter the earthworms' bodies have been described. Celomic fluid has been documented to contain various antimicrobial factors, such as lysozyme (Çotuk and Dales, 1984; Joskova *et al.*, 2009) and antimicrobial peptides (Wang *et al.*, 2003; Liu *et al.*, 2004; Li *et al.*, 2011). Among the factors that are involved in humoral immunity, particular interest has been devoted to the cytolytic components that are secreted by celomocytes. The cytolytic activity of the celomic fluid was originally demonstrated on vertebrate erythrocytes and the resulting effect was described as hemolysis. The majority of identified hemolysins exhibit broad spectra of antibacterial and/or bacteriostatic activities against pathogenic soil bacteria (Roch *et al.*, 1991; Milochau *et al.*, 1997; Eue *et al.*, 1998). Various types of pollutants, such as metallic compounds (Brulle *et al.*, 2008; Mo *et al.*, 2012) and TiO₂ nanoparticles (Bigorgne *et al.*, 2012), have been described to influence the expression of these molecules and therefore cause inappropriate immune response to invading pathogens. Earthworm calreticulin is a highly conserved calcium-binding protein that has also been shown to be affected by the presence of various pollutants in soils (Chen *et al.*, 2011; Roubalova *et al.*, 2014). It participates in the regulation of Ca²⁺ homeostasis, acts as a chaperone and is involved in the regulation of cell signaling (Wang *et al.*, 2012). It also plays a role in the stress response and immune reactions (Goo *et*

Table 2 List of pollutants that affect the activity and gene transcription of antioxidant enzymes

Pollutants that affect activities of antioxidant enzymes			
Tested species	Type of pollutant	Enzymes affected by pollutants	Reference
<i>E. fetida</i>	decabromodiphenyl ether	SOD, CAT, POD	(Zhang <i>et al.</i> , 2014)
<i>E. fetida</i>	phenanthrene	SOD, CAT, POD	(Shi <i>et al.</i> , 2013)
<i>E. fetida</i>	multi-metal-contaminated soil (Cd, Cr, Cu, Ni, Pb, and Zn)	SOD	(Zheng <i>et al.</i> , 2013)
<i>E. fetida</i>	phenanthrene, pyrene	SOD, CAT	(Wu <i>et al.</i> , 2012b)
<i>E. fetida</i>	chlortetracycline	SOD, CAT	(Lin <i>et al.</i> , 2012)
<i>E. fetida</i>	ZnO nanoparticles	SOD	(Li <i>et al.</i> , 2011)
Pollutants that affect gene expression of antioxidant enzymes			
Tested species	Type of pollutant	Genes affected by pollutants	Reference
<i>E. fetida</i>	naphthenic acids	SOD, CAT	(Wang <i>et al.</i> , 2015b)
<i>E. fetida</i>	2,2',4,4'-tetrabromodiphenyl ether	SOD, CAT	(Xu <i>et al.</i> , 2015)
<i>E. fetida</i>	copper sulphate (CuSO ₄)	SOD, CAT	(Xiong <i>et al.</i> , 2014)
<i>E. fetida</i>	silver nanoparticles	SOD, CAT	(Hayashi <i>et al.</i> , 2013)
<i>E. fetida</i>	zinc oxide (ZnO)	SOD, CAT	(Xiong <i>et al.</i> , 2012)
<i>E. fetida</i>	galaxolide, tonalide	SOD, CAT	(Chen <i>et al.</i> , 2011)

al., 2005; Kuraishi *et al.*, 2007; Silerova *et al.*, 2007; Gold *et al.*, 2010).

Enzymes involved in oxidative stress

Aerobic organisms developed efficient antioxidant defense system to protect themselves against reactive oxygen species (ROS). The major source of intracellular ROS is the mitochondrial respiratory chain (Han *et al.*, 2001; Ott *et al.*, 2007), and these radicals are also produced in smaller amounts in other cell compartments, such as the endoplasmic reticulum, the plasma and nuclear membranes, and by some oxidases (Mittler *et al.*, 2004; del Rio *et al.*, 2006; Navrot *et al.*, 2007). Free radicals were described to have an important role in cell signaling (Mates *et al.*, 2002; Scandalios, 2005; Mates *et al.*, 2008) and protection against invading pathogens (Babior *et al.*, 1975; Rossi *et al.*, 1985; Nacarelli and Fuller-Espie, 2011). Oxidative stress induces DNA modifications (Bohr, 2002), direct oxidation and inactivation of iron-sulfur (Fe-S) proteins (Fridovich, 1997), lipid peroxidation (Arai, 2014), and apoptotic events by means of caspase dependent pathways (Bearoff and Fuller-Espie, 2011). Under stressful conditions (*e.g.*, exposures to UV radiation, organic and inorganic contaminants, extreme temperatures and biotic stress), the concentrations of ROS increase, resulting in the development of oxidative stress and subsequent damage to cellular structures (Foyer and Noctor, 2005; Gill and Tuteja, 2010; Tumminello and Fuller-Espie, 2013). Antioxidant enzymes are considered

to be a primary defense that protects biological macromolecules from oxidative damage. Three groups of these enzymes play significant roles in protecting cells from oxidant stress, *i.e.*, superoxide dismutases (SODs), catalase (CAT) and peroxidases (PODs) (Mates, 2000). SODs are a ubiquitous family of metal-containing enzymes that depend on bound manganese (mitochondrial SOD), copper or zinc (intra- and extra-cellular SODs) for their antioxidant activity. SODs efficiently catalyze the dismutation of superoxide anions into hydrogen peroxide, which is substantially less toxic than superoxide, and oxygen. CAT and PODs degrade hydrogen peroxide to water. Both the enzyme activities and gene expression levels of antioxidant enzymes are frequently used to determine the effects of pollution on earthworms (Table 2).

Conclusions

This review summarized the data that have been published so far regarding the effects of various soil pollutants on the defense mechanisms of earthworms. The toxicities of these chemicals, which often enter the food chain, have been described to affect the immune system of not only invertebrates but also vertebrates, including humans. This article illustrated the various mechanisms through which the effects of pollutants are mediated on both the cellular and humoral components of the immune system. Such disruptions of the abilities of earthworms to protect

themselves against invading pathogens has been shown to be closely related to the reduced reproduction and growth rates of earthworms and increased mortality. Moreover, these immunological parameters can be used as reliable biomarkers for the detection of the pollutant-induced responses of soil organisms.

Acknowledgements

This research was supported by the Ministry of Education, Youth and Sports (CZ.1.07/2.3.00/30.0003), the Institutional Research Concept RVO 61388971 and by the project "BIOCEV - Biotechnology and Biomedicine Centre of the Academy of Sciences and Charles University in Vestec" (CZ.1.05/1.1.00/02.0109) from the European Regional Development Fund.

References

- Alves CE, Macha N, Dall Pai V. Fine structure of the epidermal cuticle of some Brazilian earthworms (Oligochaeta: Annelida). *Anat. Anz.* 155: 1-9, 1984.
- Arai H. Oxidative modification of lipoproteins. *Sub-Cell. Biochem.* 77: 103-114, 2014.
- Asensio V, Rodriguez-Ruiz A, Garmendia L, Andre J, Kille P, Morgan AJ, *et al.* Towards an integrative soil health assessment strategy: A three tier (integrative biomarker response) approach with *Eisenia fetida* applied to soils subjected to chronic metal pollution. *Sci. Total Environ.* 442: 344-365, 2013.
- Babior BM, Curnutte JT, Kipnes RS. Biological defense mechanisms. Evidence for the participation of superoxide in bacterial killing by xanthine oxidase. *J. Lab. Clin. Med.* 85: 235-244, 1975.
- Bartlett MD, Briones MJI, Neilson R, Schmidt O, Spurgeon D, Creamer RE. A critical review of current methods in earthworm ecology: From individuals to populations. *Eur. J. Soil Biol.* 46: 67-73, 2010.
- Bearoff FM, Fuller-Espie SL. Alteration of mitochondrial membrane potential ($\Delta\psi_m$) and phosphatidylserine translocation as early indicators of heavy metal-induced apoptosis in the earthworm *Eisenia hortensis*. *Inv. Surv. J.* 8: 98-108, 2011.
- Belfroid AC, Scinen W, van Gestel KC, Hermens JL, van Leeuwen KJ. Modelling the accumulation of hydrophobic organic chemicals in earthworms: Application of the equilibrium partitioning theory. *Environ. Sci. Pollut. Res. Int.* 2: 5-15, 1995.
- Beliaeff B, Burgeot T. Integrated biomarker response: a useful tool for ecological risk assessment. *Environ. Toxicol. Chem.* 21: 1316-1322, 2002.
- Belmeskine H, Haddad S, Vandelac L, Sauve S, Fournier M. Toxic effects of PCDD/Fs mixtures on *Eisenia andrei* earthworms. *Ecotox. Environ. Safe.* 80: 54-59, 2012.
- Bernaldo de Quiros IF, Benito J. Ultrastructure of gland cells associated with the chaetal follicles in the clitellar region of *Lumbricus friendi* Cognetti, 1904 (Oligochaeta). *Arch. Anat. Histol. Embryol.* 69: 91-99, 1986.
- Beschin A, Bilej M, Hanssens F, Raymakers J, Van Dyck E, Revets H, *et al.* Identification and cloning of a glucan- and lipopolysaccharide-binding protein from *Eisenia foetida* earthworm involved in the activation of prophenoloxidase cascade. *J. Biol. Chem.* 273: 24948-24954, 1998.
- Bigorgne E, Cossu-Leguille C, Bonnard M, Nahmani J. Genotoxic effects of nickel, trivalent and hexavalent chromium on the *Eisenia fetida* earthworm. *Chemosphere* 80: 1109-1112, 2010.
- Bigorgne E, Foucaud L, Caillet C, Giamberini L, Nahmani J, Thomas F, *et al.* Cellular and molecular responses of *E. fetida* coelomocytes exposed to TiO₂ nanoparticles. *J. Nanopart. Res.* 14, 2012.
- Bilej M, Rossmann P, Sinkora M, Hanusova R, Beschin A, Raes G, *et al.* Cellular expression of the cytolytic factor in earthworms *Eisenia foetida*. *Immunol. Lett.* 60: 23-29, 1998.
- Bilej M, De Baetselier P, Van Dijk E, Stijlemans B, Colige A, Beschin A. Distinct carbohydrate recognition domains of an invertebrate defense molecule recognize Gram-negative and Gram-positive bacteria. *J. Biol. Chem.* 276: 45840-45847, 2001.
- Bohr VA. Repair of oxidative DNA damage in nuclear and mitochondrial DNA, and some changes with aging in mammalian cells. *Free Radic. Biol. Med.* 32: 804-812, 2002.
- Booth LH, O'Halloran K. A comparison of biomarker responses in the earthworm *Aporrectodea caliginosa* to the organophosphorus insecticides diazinon and chlorpyrifos. *Environ. Toxicol. Chem.* 20: 2494-2502, 2001.
- Booth L, Palasz F, Darling C, Lanno R, Wickstrom M. The effect of lead-contaminated soil from Canadian prairie skeet ranges on the neutral red retention assay and fecundity in the earthworm *Eisenia fetida*. *Environ. Toxicol. Chem.* 22: 2446-2453, 2003.
- Brulle F, Cocquerelle C, Mitta G, Castric V, Douay F, Lepretre A, *et al.* Identification and expression profile of gene transcripts differentially expressed during metallic exposure in *Eisenia fetida* coelomocytes. *Dev. Comp. Immunol.* 32: 1441-1453, 2008.
- Button M, Koch I, Reimer KJ. Arsenic resistance and cycling in earthworms residing at a former gold mine in Canada. *Environ. Pollut.* 169: 74-80, 2012.
- Calisi A, Zaccarelli N, Lionetto MG, Schettino T. Integrated biomarker analysis in the earthworm *Lumbricus terrestris*: application to the monitoring of soil heavy metal pollution. *Chemosphere* 90: 2637-2644, 2013.
- Canesi L, Procházková P. The invertebrate immune system as a model for investigating the environmental impact of nanoparticles. In: Boraschi D, Duschl A (eds), *Nanoparticles and the immune system, safety and effects*, Academic Press, Oxford, pp 91-112, 2014.

- Chen C, Xue S, Zhou Q, Xie X. Multilevel ecotoxicity assessment of polycyclic musk in the earthworm *Eisenia fetida* using traditional and molecular endpoints. *Ecotoxicology* 20: 1949-1958, 2011.
- Cotelle S, Ferard JF. Comet assay in genetic ecotoxicology: A review. *Environ. Mol. Mutagen.* 34: 246-255, 1999.
- Çotuk A, Dales RP. Lysozyme activity in the coelomic fluid and coelomocytes of the earthworm *Eisenia foetida* sav. In relation to bacterial infection. *Comp. Biochem. Physiol.* 78A: 469-474, 1984.
- del Rio LA, Sandalio LM, Corpas FJ, Palma JM, Barroso JB. Reactive oxygen species and reactive nitrogen species in peroxisomes. Production, scavenging, and role in cell signaling. *Plant Physiol.* 141: 330-335, 2006.
- Drake HL, Horn MA. As the worm turns: the earthworm gut as a transient habitat for soil microbial biomes. *Ann. Rev. Microbiol.* 61: 169-189, 2007.
- Du L, Li G, Liu M, Li Y, Yin S, Zhao J, *et al.* Evaluation of DNA damage and antioxidant system induced by di-n-butyl phthalates exposure in earthworms (*Eisenia fetida*). *Ecotoxicol. Environ. Saf.* 115: 75-82, 2015.
- Duan X, Xu L, Song J, Jiao J, Liu M, Hu F, *et al.* Effects of benzo[a]pyrene on growth, the antioxidant system, and DNA damage in earthworms (*Eisenia fetida*) in 2 different soil types under laboratory conditions. *Environ. Toxicol. Chem.* 34: 283-290, 2015.
- Edwards CA. Earthworm ecology, CRC Press, Boca Raton, 2004.
- Eue I, Kauschke E, Mohrig W, Cooper EL. Isolation and characterization of earthworm hemolysins and agglutinins. *Dev. Comp. Immunol.* 22: 13-25, 1998.
- Fairbairn DW, Olive PL, O'Neill KL. The comet assay: a comprehensive review. *Mutat. Res.* 339: 37-59, 1995.
- Faust F, Kassie F, Knasmüller S, Boedecker RH, Mann M, Mersch-Sundermann V. The use of the alkaline comet assay with lymphocytes in human biomonitoring studies. *Mutat. Res.* 566: 209-229, 2004.
- Fournier M, Cyr D, Blakley B, Boermans H, Brousseau P. Phagocytosis as a biomarker of immunotoxicity in wildlife species exposed to environmental xenobiotics. *Am. Zool.* 40: 412-420, 2000.
- Foyer CH, Noctor G. Redox homeostasis and antioxidant signaling: a metabolic interface between stress perception and physiological responses. *Plant Cell* 17: 1866-1875, 2005.
- Fridovich I. Superoxide anion radical (O₂⁻), superoxide dismutases, and related matters. *J. Biol. Chem.* 272: 18515-18517, 1997.
- Fugere N, Brousseau P, Krzystyniak K, Coderre D, Fournier M. Heavy metal-specific inhibition of phagocytosis and different in vitro sensitivity of heterogeneous coelomocytes from *Lumbricus terrestris* (Oligochaeta). *Toxicology* 109: 157-166, 1996.
- Gill SS, Tuteja N. Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant physiology and biochemistry : PPB/ Societe francaise de physiologie vegetale* 48: 909-930, 2010.
- Gold LI, Eggleton P, Sweetwyne MT, Van Duyn LB, Greives MR, Naylor SM, *et al.* Calreticulin: non-endoplasmic reticulum functions in physiology and disease. *FASEB. J.* 24: 665-683, 2010.
- Goo TW, Park S, Jin BR, Yun EY, Kim I, Nho SK, *et al.* Endoplasmic reticulum stress response of *Bombyx mori* calreticulin. *Mol. Biol. Rep.* 32: 133-139, 2005.
- Han D, Williams E, Cadenas E. Mitochondrial respiratory chain-dependent generation of superoxide anion and its release into the intermembrane space. *Biochem. J.* 353: 411-416, 2001.
- Han Y, Zhu L, Wang J, Wang J, Xie H, Zhang S. Integrated assessment of oxidative stress and DNA damage in earthworms (*Eisenia fetida*) exposed to azoxystrobin. *Ecotoxicol. Environ. Saf.* 107: 214-219, 2014.
- Hankard PK, Svendsen C, Wright J, Wienberg C, Fishwick SK, Spurgeon DJ, *et al.* Biological assessment of contaminated land using earthworm biomarkers in support of chemical analysis. *Sci. Total. Environ.* 330: 9-20, 2004.
- Hayashi Y, Engelmann P, Foldbjerg R, Szabo M, Somogyi I, Pollak E, *et al.* Earthworms and humans in vitro: Characterizing evolutionarily conserved stress and immune responses to silver nanoparticles. *Environ. Sci. Technol.* 46: 4166-4173, 2012.
- Hayashi Y, Heckmann LH, Simonsen V, Scott-Fordsmand JJ. Time-course profiling of molecular stress responses to silver nanoparticles in the earthworm *Eisenia fetida*. *Ecotoxicol. Environ. Saf.* 98: 219-226, 2013.
- ISO. Soil quality - Effects of pollutants on earthworms (*Eisenia fetida*). Part 1: Determination of acute toxicity using artificial soil substrate International Organisation for Standardization, Geneva, Switzerland, 1993.
- ISO. Soil quality - Effects of pollutants on earthworms (*Eisenia fetida*). Part 2: Determination of effects on reproduction. International Organisation for Standardization, Geneva, Switzerland, 1998.
- Jager T, Fleuren RH, Hogendoorn EA, de Korte G. Elucidating the routes of exposure for organic chemicals in the earthworm, *Eisenia andrei* (Oligochaeta). *Environ. Sci. Technol.* 37: 3399-3404, 2003.
- Jager T, van der Wal L, Fleuren RH, Barendregt A, Hermens JL. Bioaccumulation of organic chemicals in contaminated soils: evaluation of bioassays with earthworms. *Environ. Sci. Technol.* 39: 293-298, 2005.
- Joskova R, Silerova M, Prochazkova P, Bilej M. Identification and cloning of an invertebrate-type lysozyme from *Eisenia andrei*. *Dev. Comp. Immunol.* 33: 932-938, 2009.
- Kammenga JE, Dallinger R, Donker MH, Kohler HR, Simonsen V, Triebkorn R., *et al.* Biomarkers in terrestrial invertebrates for ecotoxicological soil

- risk assessment. *Rev. Environ. Contam. Toxicol.* 164: 93-147, 2000.
- Klobucar GI, Stambuk A, Srut M, Husnjak I, Merkas M, Traven L, *et al.* *Aporrectodea caliginosa*, a suitable earthworm species for field based genotoxicity assessment? *Environ. Pollut.* 159: 841-849, 2011.
- Kuraishi T, Manaka J, Kono M, Ishii H, Yamamoto N, Koizumi K, *et al.* Identification of calreticulin as a marker for phagocytosis of apoptotic cells in *Drosophila*. *Exp. Cell Res.* 313: 500-510, 2007.
- Lee KE. Earthworms: Their ecology and relationships with soil and land use. Academic Press, Sydney, 1985.
- Li LZ, Zhou DM, Peijnenburg WJ, van Gestel CA, Jin SY, Wang YJ, *et al.* Toxicity of zinc oxide nanoparticles in the earthworm, *Eisenia fetida* and subcellular fractionation of Zn. *Environ. Int.* 37: 1098-1104, 2011.
- Li M, Liu Z, Xu Y, Cui Y, Li D, Kong Z. Comparative effects of Cd and Pb on biochemical response and DNA damage in the earthworm *Eisenia fetida* (Annelida, Oligochaeta). *Chemosphere* 74: 621-625, 2009.
- Li WL, Li SS, Zhong J, Zhu Z, Liu JZ, Wang WH. A novel antimicrobial peptide from skin secretions of the earthworm, *Pheretima guillelmi* (Michaelson). *Peptides* 32: 1146-1150, 2011.
- Lin D, Zhou Q, Xu Y, Chen C, Li Y. Physiological and molecular responses of the earthworm (*Eisenia fetida*) to soil chlortetracycline contamination. *Environ. Pollut.* 171: 46-51, 2012.
- Lin D, Li Y, Zhou Q, Xu Y, Wang D. Effect of triclosan on reproduction, DNA damage and heat shock protein gene expression of the earthworm *Eisenia fetida*. *Ecotoxicology* 23: 1826-1832, 2014.
- Liu T, Zhu LS, Han YN, Wang JH, Wang J, Zhao Y. The cytotoxic and genotoxic effects of metalaxyl-M on earthworms (*Eisenia fetida*). *Environ. Toxicol. Chem.* 33: 2344-2350, 2014.
- Liu Y, Zhou Q, Xie X, Lin D, Dong L. Oxidative stress and DNA damage in the earthworm *Eisenia fetida* induced by toluene, ethylbenzene and xylene. *Ecotoxicology* 19: 1551-1559, 2010.
- Liu YQ, Sun ZJ, Wang C, Li SJ, Liu YZ. Purification of a novel antibacterial short peptide in earthworm *Eisenia foetida*. *Acta Biochim. Biophys. Sin. (Shanghai)* 36: 297-302, 2004.
- Lourenco JI, Pereira RO, Silva AC, Morgado JM, Carvalho FP, Oliveira JM, *et al.* Genotoxic endpoints in the earthworms sub-lethal assay to evaluate natural soils contaminated by metals and radionuclides. *J. Hazard. Mat.* 186: 788-795, 2011.
- Maboeta M, Reinecke SA, Reinecke AJ. Linking lysosomal biomarker and population responses in a field population of *Aporrectodea caliginosa* (Oligochaeta) exposed to the fungicide copper oxychloride. *Ecotox. Environ. Saf.* 56: 411-418, 2003.
- Mates JM. Effects of antioxidant enzymes in the molecular control of reactive oxygen species toxicology. *Toxicology* 153: 83-104, 2000.
- Mates JM, Perez-Gomez C, Nunez de Castro I, Asenjo M, Marquez J. Glutamine and its relationship with intracellular redox status, oxidative stress and cell proliferation/death. *Int. J. Biochem. Cell Biol.* 34: 439-458, 2002.
- Mates JM, Segura JA, Alonso FJ, Marquez J. Intracellular redox status and oxidative stress: implications for cell proliferation, apoptosis, and carcinogenesis. *Archiv. Toxicol.* 82: 273-299, 2008.
- Milochau A, Lassegues M, Valembois P. Purification, characterization and activities of two hemolytic and antibacterial proteins from coelomic fluid of the annelid *Eisenia fetida andrei*. *Biochim. Biophys. Acta* 1337: 123-132, 1997.
- Mittler R, Vanderauwera S, Gollery M, Van Breusegem F. Reactive oxygen gene network of plants. *Trends Plant Sci.* 9: 490-498, 2004.
- Mo XH, Qiao YH, Sun ZJ, Sun XF, Li Y. Molecular toxicity of earthworms induced by cadmium contaminated soil and biomarkers screening. *J. Environ. Sci.-China* 24: 1504-1510, 2012.
- Moore MN. Cellular-responses to pollutants. *Mar. Pollut. Bull.* 16: 134-139, 1985.
- Moore MN. Lysosomal cytochemistry in marine environmental monitoring. *Histochem. J.* 22: 187-191, 1990.
- Nacarelli T, Fuller-Espie SL. Pathogen-associated molecular pattern-induced mitochondrial membrane depolarization in the earthworm *Eisenia hortensis*. *J. Invertebr. Pathol.* 108: 174-179, 2011.
- Nahmani J, Hodson ME, Black S. A review of studies performed to assess metal uptake by earthworms. *Environ. Pollut.* 145: 402-424, 2007.
- Navrot N, Rouhier N, Gelhaye E, Jacquot JP. Reactive oxygen species generation and antioxidant systems in plant mitochondria. *Physiol. Plantarum* 129: 185-195, 2007.
- OECD. Guideline for the testing of chemicals. No. 207, Earthworm, acute toxicity tests. Organisation for Economic Cooperation and Development, Paris, France, 1984.
- OECD. Guideline for the testing of chemicals. No. 222, Earthworm reproduction test (*Eisenia fetida/Eisenia andrei*). Organisation for Economic Cooperation and Development, Paris, France, 2004.
- Otomo PV, Reinecke SA, Reinecke AJ. Using the comet assay to assess the combined and separate genotoxic effects of Cd and Zn in *Eisenia andrei* (Oligochaeta) at different temperatures. *Bull. Environ. Contam. Tox.* 92: 285-288, 2014.
- Ott M, Gogvadze V, Orrenius S, Zhivotovsky B. Mitochondria, oxidative stress and cell death. *Apoptosis* 12: 913-922, 2007.
- Owen J, Hedley BA, Svendsen C, Wren J, Jonker MJ, Hankard PK, *et al.* Transcriptome profiling of developmental and xenobiotic responses in a keystone soil animal, the oligochaete annelid *Lumbricus rubellus*. *BMC Genomics* 9: 266, 2008.

- Pakarinen K, Petersen EJ, Leppanen MT, Akkanen J, Kukkonen JV. Adverse effects of fullerenes (nC60) spiked to sediments on *Lumbricus variegatus* (Oligochaeta). *Environ. Pollut.* 159: 3750-3756, 2011.
- Patel M, Francis J, Cooper EL, Fuller-Espie SL. Development of a flow cytometric, non-radioactive cytotoxicity assay in *Eisenia fetida*: An in vitro system designed to analyze immunosuppression of natural killer-like coelomocytes in response to 7,12 dimethylbenz[a]anthracene (DMBA). *Eur. J. Soil Biol.* 43: S97-S103, 2007.
- Plytycz B, Lis-Molenda U, Cygal M, Kielbasa E, Grebosz A, Duchnowski M, et al. Riboflavin content of coelomocytes in earthworm (*Dendrodrilus rubidus*) field populations as a molecular biomarker of soil metal pollution. *Environ. Pollut.* 157: 3042-3050, 2009.
- Rahemtulla F, Lovtrup S. The comparative biochemistry of invertebrate mucopolysaccharides II. Nematoda; Annelida. *Comp. Biochem. Physiol.* 49B: 639-646, 1974.
- Roch P, Lassegues M, Valembos P. Antibacterial activity of *Eisenia fetida andrei* coelomic fluid: III. Relationship within the polymorphic hemolysins. *Dev. Comp. Immunol.* 15: 27-32, 1991.
- Rossi F, Della Bianca V, de Togni P. Mechanisms and functions of the oxygen radicals producing respiration of phagocytes. *Comp. Immunol. Microbiol. Infect. Dis.* 8: 187-204, 1985.
- Roubalova R, Dvorak J, Prochazkova P, Elhottova D, Rossmann P, Skanta F, et al. The effect of dibenzo-p-dioxin- and dibenzofuran-contaminated soil on the earthworm *Eisenia andrei*. *Environ. Pollut.* 193: 22-28, 2014.
- Ruiz E, Alonso-Azcarate J, Rodriguez L. *Lumbricus terrestris* L. activity increases the availability of metals and their accumulation in maize and barley. *Environ. Pollut.* 159: 722-728, 2011.
- Sanchez-Hernandez JC, Narvaez C, Sabat P, Martinez Mocillo S. Integrated biomarker analysis of chlorpyrifos metabolism and toxicity in the earthworm *Aporrectodea caliginosa*. *Sci. Total Environ.* 490: 445-455, 2014.
- Sauve S, Hendawi M, Brousseau P, Fournier M. Phagocytic response of terrestrial and aquatic invertebrates following in vitro exposure to trace elements. *Ecotox. Environ. Safe.* 52: 21-29, 2002.
- Scandalios JG. Oxidative stress: molecular perception and transduction of signals triggering antioxidant gene defenses. *Braz. J. Med. Biol. Res.* 38: 995-1014, 2005.
- Sforzini S, Boeri M, Dagnino A, Oliveri L, Bolognesi C, Viarengo A. Genotoxicity assessment in *Eisenia andrei* coelomocytes: A study of the induction of DNA damage and micronuclei in earthworms exposed to B[a]P- and TCDD-spiked soils. *Mutat. Res.-Gen. Tox. En.* 746: 35-41, 2012.
- Sforzini S, Moore MN, Boeri M, Bencivenga M, Viarengo A. Effects of PAHs and dioxins on the earthworm *Eisenia andrei*: a multivariate approach for biomarker interpretation. *Environ. Pollut.* 196: 60-71, 2015.
- Shi Z, Xu L, Wang N, Zhang W, Li H, Hu F. Pseudo-basal levels of and distribution of anti-oxidant enzyme biomarkers in *Eisenia fetida* and effect of exposure to phenanthrene. *Ecotoxicol. Environ. Saf.* 95: 33-38, 2013.
- Silerova M, Kauschke E, Prochazkova P, Joskova R, Tuckova L, Bilej M. Characterization, molecular cloning and localization of calreticulin in *Eisenia fetida* earthworms. *Gene* 397: 169-177, 2007.
- Sima P. Annelid coelomocytes and haemocytes: roles in cellular immune reactions. In: Vetvicka V, Sima P, Cooper EL, Bilej M, Roch P (eds), *Immunology of Annelids*, CRC Press, Boca Raton, Ann Arbor, London, Tokyo, pp 11-165, 1994.
- Singh NP, McCoy MT, Tice RR, Schneider EL. A simple technique for quantitation of low levels of DNA damage in individual cells. *Exp. Cell Res.* 175: 184-191, 1988.
- Skanta F, Roubalova R, Dvorak J, Prochazkova P, Bilej M. Molecular cloning and expression of TLR in the *Eisenia andrei* earthworm. *Dev. Comp. Immunol.* 41: 694-702, 2013.
- Soderhall K, Cerenius L. Role of the prophenoloxidase-activating system in invertebrate immunity. *Curr. Opin. Immunol.* 10: 23-28, 1998.
- Spurgeon DJ, Ricketts H, Svendsen C, Morgan AJ, Kille P. Hierarchical responses of soil invertebrates (earthworms) to toxic metal stress. *Environ. Sci. Technol.* 39: 5327-5334
- Suzuki MM, Cooper EL, Eyambe GS, Goven AJ, Fitzpatrick LC, Venables BJ. Polychlorinated-biphenyls (Pcbs) depress allogeneic natural cytotoxicity by earthworm coelomocytes. *Environ. Toxicol. Chem.* 14: 1697-1700, 1995.
- Suzuki MM, Cooper EL. Spontaneous cytotoxic earthworm leukocytes kill K562 tumor cells. *Zool. Sci.* 12:443-451, 1995.
- Tumminello RA, Fuller-Espie SL. Heat stress induces ROS production and histone phosphorylation in coelomocytes of *Eisenia hortensis*. *Inv. Surv. J.* 10: 50-57, 2013.
- Valembos P, Roch P, Lassegues M. Antibacterial molecules in annelids. In: Brehelin M (ed.), *Immunity in invertebrates*, Springer-Verlag, Berlin, Heidelberg, New York, Tokyo, pp 74-93, 1986.
- Van Der Ploeg MJ, Handy RD, Heckmann LH, Van Der Hout A, Van Den Brink NW. C60 exposure induced tissue damage and gene expression alterations in the earthworm *Lumbricus rubellus*. *Nanotoxicology* 7: 432-440, 2013.
- Venkateswara Rao J, Kavitha P, Padmanabha Rao A. Comparative toxicity of tetra ethyl lead and lead oxide to earthworms, *Eisenia fetida* (Savigny). *Environ. Res.* 92: 271-276, 2003.
- Ville P, Roch P, Cooper EL, Masson P, Narbonne JF. Pcbs increase molecular-related activities (lysozyme, antibacterial, hemolysis, proteases) but inhibit macrophage-related functions (phagocytosis, wound-healing) in earthworms. *J. Invertebr. Pathol.* 65: 217-224, 1995.
- Wang J, Cao X, Chai L, Liao J, Huang Y, Tang X. Oxidative damage of naphthenic acids on the

- Eisenia fetida* earthworm. Environ. Toxicol. doi: 10.1002/tox.22139, 2015a.
- Wang J, Cao X, Sun J, Chai L, Huang Y, Tang X. Transcriptional responses of earthworm (*Eisenia fetida*) exposed to naphthenic acids in soil. Environ. Pollut. 204: 264-270, 2015b.
- Wang WA, Groenedyk J, Michalak M. Calreticulin signalling in health and disease. Int. J. Biochem. Cell. Biol. 44: 842-846, 2012.
- Wang X, Wang X, Zhang Y, Qu X, Yang S. An antimicrobial peptide of the earthworm *Pheretima tschiliensis*: cDNA cloning, expression and immunolocalization. Biotechnol. Lett. 25: 1317-1323, 2003.
- Weeks JM, Svendsen C. Neutral red retention by lysosomes from earthworm (*Lumbricus rubellus*) coelomocytes: A simple biomarker of exposure to soil copper. Environ. Toxicol. Chem. 15: 1801-1805, 1996.
- Wen B, Liu Y, Hu XY, Shan XQ. Effect of earthworms (*Eisenia fetida*) on the fractionation and bioavailability of rare earth elements in nine Chinese soils. Chemosphere 63: 1179-1186, 2006.
- Wu SJ, Zhang HX, Hu Y, Li HL, Chen JM. Effects of 1,2,4-trichlorobenzene on the enzyme activities and ultrastructure of earthworm *Eisenia fetida*. Ecotox. Environ. Safe. 76: 175-181, 2012a.
- Wu S, Zhang H, Zhao S, Wang J, Li H, Chen J. Biomarker responses of earthworms (*Eisenia fetida*) exposed to phenanthrene and pyrene both singly and combined in microcosms. Chemosphere 87: 285-293, 2012b.
- Xiong W, Bai L, Muhammad RU, Zou M, Sun Y. Molecular cloning, characterization of copper/zinc superoxide dismutase and expression analysis of stress-responsive genes from *Eisenia fetida* against dietary zinc oxide. Comp. Biochem. Physiol. 155C: 416-422, 2012.
- Xiong W, Ding X, Zhang Y, Sun Y. Ecotoxicological effects of a veterinary food additive, copper sulphate, on antioxidant enzymes and mRNA expression in earthworms. Environ. Toxicol. Pharmacol. 37: 134-140, 2014.
- Xu XB, Shi YJ, Lu YL, Zheng XQ, Ritchie RJ. Growth inhibition and altered gene transcript levels in earthworms (*Eisenia fetida*) exposed to 2,2',4,4'-tetrabromodiphenyl ether. Arch. Environ. Contam. Toxicol. 69: 1-7, 2015.
- Zhang W, Liu K, Chen L, Chen L, Lin K, Fu R. A multi-biomarker risk assessment of the impact of brominated flame retardant-decabromodiphenyl ether (BDE209) on the antioxidant system of earthworm *Eisenia fetida*. Environ. Toxicol. Pharmacol. 38: 297-304, 2014.
- Zheng K, Liu Z, Li Y, Cui Y, Li M. Toxicological responses of earthworm (*Eisenia fetida*) exposed to metal-contaminated soils. Environ. Sci. Pollut. Res. Int. 20: 8382-8390, 2013.